Acta Crystallographica Section E

Structure Reports Online

ISSN 1600-5368

Suchada Chantrapromma, a* Nawong Boonnak, a Hoong-Kun Fun,b‡ Shazia Anjumc and Atta-ur-Rahman^c

^aDepartment of Chemistry, Faculty of Science, Prince of Songkla University, Hat-Yai, Songkhla 90112, Thailand, bX-ray Crystallography Unit, School of Physics, Universiti Sains Malaysia, 11800 USM, Penang, Malaysia, and CHEJ Research Institute of Chemistry, International Center for Chemical Sciences, University of Karachi, Karachi 75270, Pakistan

‡ Additional correspondence author, email: hkfun@usm.mv

Correspondence e-mail: suchada.c@psu.ac.th

Key indicators

Single-crystal X-ray study T = 293 KMean $\sigma(C-C) = 0.006 \text{ Å}$ R factor = 0.087 wR factor = 0.230 Data-to-parameter ratio = 13.4

For details of how these key indicators were automatically derived from the article, see http://journals.iucr.org/e.

5,9,10-Trihydroxy-2,2-dimethyl-12-(3-methylbut-2-enyl)-2H,6H-pyrano[3,2-b]xanthen-6-one methanol solvate

The title compound, C23H22O6·CH3OH, has an essentially planar xanthone nucleus fused to a chromene ring which adopts a sofa conformation. The 3-methylbut-2-enyl substituent is axially attached to the xanthone ring system, with a (+)-anticlinal conformation. The crystal structure is stabilized by O-H···O and C-H···O intramolecular and intermolecular interactions.

Received 31 May 2005 Accepted 8 June 2005 Online 17 June 2005

Comment

Cratoxylum is a small genus belonging to the Guttiferae family, with at least six species distributed in several Southeast Asian countries (Iinuma et al., 1996). Species of this genus have been used for their diuretic, stomachic, and tonic effects (Kitanov et al., 1988), as well as for diarrhea and flatulence (Anderson, 1986), and for food poisoning and internal bleeding (Grosvenor et al., 1995). Some species of this genus exhibit antimalarial and antiprotozoal activity, and are slightly cytoxic against human L6 cells (Seo et al., 2002; Zakaria, 2004). In our continuing search for bioactive compounds from Thai medicinal plants (Chantrapromma et al., 2003; Boonnak et al., 2005), we have investigated Cratoxylum formosum ssp. pruniflorum, a shrub which is known locally to Thais as Tuikhon. Tuikhon was collected from Nhongkhai province and is widely distributed in the north-eastern part of Thailand. We have isolated the title compound, (I), xanthone V₁, for the first time from Cratoxylum formosum ssp. pruniflorum. It was previously isolated from Vismia guineensis (Botta et al., 1986) and Garcinia latissima (Ito et al., 1997).

Because of the puckering of atoms O6 and C16, the title molecule is chiral, but crystallized in the centrosymmetric space group $P2_1/c$. This indicates that the crude extract from which the compound was obtained is a racemic mixture and that (I) was produced by non-enzymatic cyclization of a side chain. A closely related structure, having an identical skeleton, is dulxanthone E (Kosela et al., 1999).

© 2005 International Union of Crystallography Printed in Great Britain - all rights reserved

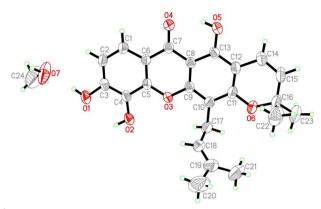


Figure 1
The structure of the title compound, showing 50% probability displacement ellipsoids and the atom-numbering scheme.

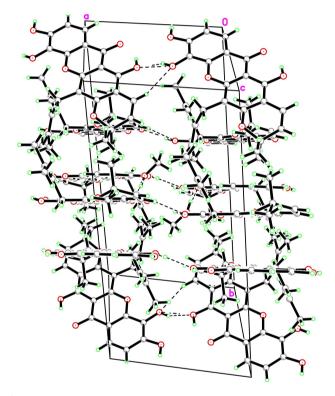


Figure 2
A view of the molecular packing. Dashed lines indicate hydrogen bonds.

The xanthone skeleton (rings A, B and C) is essentially planar, with a maximum deviation of 0.044 (2) Å for atom O3. The chromene ring D is in a sofa conformation, with puckering parameter Q=0.297 (5) Å (Cremer & Pople, 1975), atom C16 having the maximum deviation of 0.190 (4) Å. The two methyl groups are axially and bisectionally attached to the chromene ring at atom C16, with torsion angles C14—C15—C16—C22 of -93.7 (6)° and C14—C15—C16—C23 of 141.8 (5)°. The 3-methylbut-2-enyl substituent is attached to ring C at C10, with C11—C10—C17—C18 = 97.4 (4)°, indicating a (+)-anticlinal conformation (Fig. 1).

The bond lengths and angles in (I) have normal values (Allen *et al.*, 1987) and are comparable to those in dulxanthone E (Kosela *et al.*, 1999). Intramolecular and inter-

molecular $O-H\cdots O$ and weak $C-H\cdots O$ interactions are observed (Table 2). The molecules are linked together by these interactions to form a three-dimensional molecular network (Fig. 2).

Experimental

Air-dried barks of *C. formosum* ssp. *pruniflorum* (4 kg) were ground and extracted with hexane and CH_2Cl_2 (2 × 20 l for each solvent) for 5 d at room temperature. The residue obtained after evaporation of the solvent was subjected to quick column chromatography (QCC) over silica gel and eluted with a gradient of EtOAc-hexane to afford ten fractions (F1–F10). Fraction F6 (3.72 g) was separated by column chromatography (CC), eluted with 15% EtOAc-hexane to afford seven fractions (6 A–6 G). Fraction 6B was further purified by CC with 30% EtOAc-hexane to give two fractions (6BA and 6BB). Fraction 6BA was recrystallized from $CHCl_3$ – CH_3OH (8:2 ν/ν) to give brown single crystals of (I) after several days (m.p. 491–492 K).

Crystal data

$C_{23}H_{22}O_6\cdot CH_4O$	$D_x = 1.286 \text{ Mg m}^{-3}$
$M_r = 426.45$	Mo $K\alpha$ radiation
Monoclinic, $P2_1/c$	Cell parameters from 11187
a = 9.9258 (11) Å	reflections
b = 20.089 (2) Å	$\theta = 2.0 – 25.0^{\circ}$
c = 11.8605 (13) Å	$\mu = 0.09 \text{ mm}^{-1}$
$\beta = 111.324 (2)^{\circ}$ $V = 2203.1 (4) \text{ Å}^3$	T = 293 (2) K
$V = 2203.1 \text{ (4) Å}^3$	Block, brown
Z = 4	$0.66\times0.29\times0.13~\text{mm}$
Data collection	
Siemens SMART CCD area-	3875 independent reflections

Siemens SMART CCD areadetector diffractometer ω scans Absorption correction: multi-scan (SADABS; Sheldrick, 1996) $T_{\min} = 0.968$, $T_{\max} = 0.988$

11187 measured reflections

3309 reflections with $I > 2\sigma(I)$ $R_{\text{int}} = 0.023$ $\theta_{\text{max}} = 25.0^{\circ}$ $h = -11 \rightarrow 11$ $k = -23 \rightarrow 23$ $l = -14 \rightarrow 6$

Refinement

_	
Refinement on F^2	$w = 1/[\sigma^2(F_0^2) + (0.0696P)^2]$
$R[F^2 > 2\sigma(F^2)] = 0.087$	+ 5.0192 <i>P</i>]
$wR(F^2) = 0.230$	where $P = (F_0^2 + 2F_c^2)/3$
S = 1.11	$(\Delta/\sigma)_{\rm max} < 0.001$
3875 reflections	$\Delta \rho_{\text{max}} = 0.25 \text{ e Å}^{-3}$
289 parameters	$\Delta \rho_{\min} = -0.26 \text{ e Å}^{-3}$
H-atom parameters constrained	

Table 1Selected geometric parameters (Å, °).

= =			
O1-C3	1.356 (4)	O5-C13	1.344 (5)
O2-C4	1.354 (5)	O6-C11	1.358 (5)
O3-C5	1.365 (4)	O6-C16	1.468 (5)
O3-C9	1.372 (4)	O7-C24	1.369 (8)
O4-C7	1.252 (4)	C18-C19	1.314 (7)
C5-O3-C9	119.3 (3)	O6-C16-C23	103.9 (4)
C11-O6-C16	119.1 (3)	C15-C16-C23	112.0 (4)
C9-C10-C17	121.6 (3)	O6-C16-C22	107.4 (4)
C11-C10-C17	122.4 (3)	C15-C16-C22	111.4 (5)
C14-C15-C16-C23	141.8 (5)	C11-C10-C17-C18	97.4 (4)
C14-C15-C16-C22	-93.7(6)	C17-C18-C19-C20	-178.8(7)
C9-C10-C17-C18	-81.4(5)	C17-C18-C19-C21	-1.4(11)

organic papers

Table 2 Hydrogen-bond geometry (Å, °).

$D-H\cdots A$	D-H	$H \cdot \cdot \cdot A$	$D \cdot \cdot \cdot A$	D $ H$ $\cdot \cdot \cdot A$
O1−H1 <i>A</i> ···O7 ⁱ	0.82	1.85	2.668 (5)	171
$O2-H2A\cdots O1^{i}$	0.82	2.31	2.718 (5)	111
$O2-H2A\cdots O5^{ii}$	0.82	2.01	2.792 (5)	160
$O5-H5A\cdots O4^{i}$	0.82	1.84	2.570 (4)	148
$O7-H7A\cdots O4^{iii}$	0.82	1.97	2.789 (4)	171
$C14-H14A\cdots O2^{iv}$	0.93	2.51	3.394 (6)	159
$C17-H17B\cdots O6^{i}$	0.97	2.42	2.807 (6)	103

Symmetry codes: (i) x, y, z; (ii) x + 1, y, z; (iii) -x, -y + 1, -z + 1; (iv) x - 1, y, z.

H atoms were placed in calculated positions with an O—H distance of 0.82 Å and C—H distances in the range 0.93–0.98 Å. The $U_{\rm iso}$ values were constrained to be $1.5U_{\rm eq}$ of the carrier atoms for hydroxyl and methyl H atoms and $1.2U_{\rm eq}$ for the other H atoms.

Data collection: *SMART* (Siemens, 1996); cell refinement: *SAINT* (Siemens, 1996); data reduction: *SAINT*; program(s) used to solve structure: *SHELXTL* (Sheldrick, 1997); program(s) used to refine structure: *SHELXTL*; molecular graphics: *SHELXTL*; software used to prepare material for publication: *SHELXTL* and *PLATON* (Spek, 2003).

NB thanks the Development and Promotion of Science and Technology Talents Project and the PSU Graduate Research Fund for partial financial support. The authors thank Prince of Songkla University, the Pakistan Government and also the Malaysian Government and Universiti Sains Malaysia for the Scientific Advancement Grant Allocation (SAGA) grant No. 304/PFIZIK/635003/A118.

References

Allen, F. H., Kennard, O., Watson, D. G., Brammer, L., Orpen, A. G. & Taylor, R. (1987). J. Chem. Soc. Perkin Trans. 2, pp. S1–19.

Anderson, E. F. (1986). Econ. Bot. 40, 442-450.

Botta, B., Monache, G. D., Monache, F. D., Mariti Bettolo, G. B. & Menichini, F. (1986). *Phytochemistry*, **25**, 1217–1219.

Boonnak, N., Chantrapromma, S., Fun, H.-K., Anjum, Ali, S., Rahman, A. & Karalai, C. (2005). *Acta Cryst.* E**61**, o410–o412.

Chantrapromma, S., Fun, H.-K., Ibrahim A. R., Laphookhieo, S. & Karalai, C. (2003). *Acta Cryst.* E**59**, o1864–o1866.

Cremer, D. & Pople, J. A. (1975). J. Am. Chem. Soc. 97, 1354-1358.

Grosvenor, P. W., Gothard, P. K., William, N. C., Supriono, A. & Gray, D. O. (1995). J. Ethnopharmacol. 45, 75–95.

Iinuma, M., Tosa, H., Ito, T., Tanaka, T.& Madulid, D. A. (1996).
Phytochemistry, 42, 1195–1198.

Ito, C., Miyamoto, Y., Nakayama, M., Kawai, Y., Rao, K. S. & Furukawa, H. (1997). Chem. Pharm. Bull. 45, 1403–1413.

Kitanov, G. M., Assenov, I. & The Van, D. (1988). *Pharmazie*, 43, H12–H13.
Kosela, S., Hu, L. H., Yip, S. C., Rachmatia, T., Sukri, T., Daulay, T. S., Tan, G. K., Vittal, J. J. & Sim, K. Y. (1999). *Phytochemistry*, 52, 1375–1377.

Seo, E. K., Kim, N. C., Wani, M. C., Wall, M. E., Navarro, H., Burgess, J. P., Kawanishi, K., Kardono, L. B. S., Riswan, S., Rose, W. C., Fairchild, C. R., Farnsworth, N. R. & Kinghorn, D. A. (2002). J. Nat. Prod. 65, 299–305.

Sheldrick, G. M. (1996). SADABS. University of Göttingen, Germany.

Sheldrick, G. M. (1997). SHELXTL. Version 5.1. Bruker AXS Inc., Madison, Wisconsin, USA.

Siemens (1996). SMART and SAINT. Siemens Analytical X-Ray Instruments Inc., Madison, Wisconsin, USA.

Spek, A. L. (2003). J. Appl. Cryst. 36, 7-13.

Zakaria, H. (2004). Planta Med. 70, 706-710.